

Selfing revealed positive values than backcrossing for yield and yield enhancing traits among tomato segregating populations generated from *Solanum lycopersicum* × *S. pimpinellifolium* crosses under tropical humid climate

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22 **Abstract**

23 The objectives of this study were to assess phenotypic variability among F₃ and BC₁F₂ tomato
24 populations, and apply genotype by yield*trait (GYT) biplots for population and line selection
25 based on multiple traits. Four diverse cultivated parents ('CLN2498D' and 'CLN2417H' from
26 Ethiopia; 'UC Dan INDIA' and 'Tima' from Nigeria), and wild parent 'LA2093' were used to
27 generate 276 potential breeding lines. The lines were categorized into eight populations
28 ('Pop_1_W/H1', 'Pop_2_W/H2', 'Pop_3_W/D1', 'Pop_4_W/D2', 'Pop_5_W/T1',
29 'Pop_6_W/T2', 'Pop_7_W/U1', and 'Pop_8_W/U2'), and evaluated twice in the field using 19 ×
30 15 alpha-lattice design with two replicates. Significant differences were observed among lines

31 and populations for all yield enhancing traits. ‘Pop_1_W/H1’, ‘pop_4_W/D2’ and
32 ‘pop_6_W/T2’ expressed the highest genetic divergence for plant height, number of leaves, total
33 flower and fruit number, and fruit weight. GYT biplots revealed that all yield*trait interactions
34 had a positive correlation with each other. F₃ populations, ‘Pop_5_W/T1’ and ‘pop_1_W/H1’
35 exhibited the best performance for majority of the yield*trait combinations. Hierarchical
36 clustering on principal components (HCPC) revealed overlapping lines (70.58% of Cluster D
37 lines) and (54.05% of Cluster U lines) from the two F₃ populations. In BC₁F₂ population, 32.35%
38 of the 34 original lines of Cluster D and 48.48% of Cluster T lines overlapped between Clusters
39 D and T, while 18.18% of Cluster T lines and 8.82% of Cluster H lines were transgressive
40 between Clusters T and H. Transgressive segregants ‘0210U1’, ‘0211U1’, and ‘0171T1’ of
41 selfed population using multivariate analysis were believed to represent potential sources of
42 novel genetic variation for future tomato breeding.

43 **Keywords:** *Solanum pimpinellifolium*, genotype by yield*trait biplot, humid environment, multi-
44 traits, tomato population improvement, superiority index

45 **1. Introduction**

46 Tomato (*Solanum lycopersicum*) is the world’s second most important vegetable crop after
47 potato (FAOSTAT 2020). Its global production has increased by over 54% between 2000 and
48 2014 (FAO 2017). In 2019, 182 million metric tons of tomato was produced which was valued at
49 about US \$95 billion (FAOSTAT 2020). The increase in tomato fruit production hinges on the
50 use of improved and high yielding cultivars (Wang et al. 2020).

51 However, the consistent increase in world production over the years has not adequately satisfied
52 the global tomato fruit demand. Asfaw (2021) observed a supply gap in tomato fruit demand.

53 Atugwu et al. (2019) attributed this scarcity to environmental limitations placed on the most
54 available high premium cultivars by the humid environmental conditions. Tomato leaf size, fruit
55 yields and fruit quality are reduced drastically at lower vapor pressure deficit or higher humidity.
56 There is also significant increase in flower and fruit abortion, high disease occurrence and rapid
57 fruit decay both in field and after harvest under high humidity conditions (Uguru and Atugwu
58 2000).

59 The challenge in terms of yield loss, reduced productivity and susceptibility of tomato to both
60 biotic and abiotic stress factors caused by high humidity of the production environment is
61 increasingly drawing attention of the stakeholders in tomato research and production. For
62 instance, Xu (2007) reported significant yield loss in tomato grown under high air humidity,
63 which was suspected to be a result of lower photosynthesis-related activities and lower
64 photosynthetic capacity evident under such growing condition compared to tomato exposed to
65 low humidity. Alam et al. (2010) observed extreme sensitivity of tomato to hot as well as humid
66 environments, which affected yield and other physiological processes negatively.

67 In tackling new challenges, breeders have relied on wild relatives of plant for novel gene donor.
68 Among the wild relatives of tomato, *Solanum pimpinellifolium* stands out as a good gene donor.
69 This is due to its large genetic variability, high compatibility, excellent growth architecture, good
70 fruit organoleptic qualities, prolonged fruit shelf life, profuse flowering and fruiting, and high
71 adaptability to humid environment (Miller and Tanksely 1990; Foolad 2007; Rodriguez et al.
72 2006; Rodriguez et al. 2010; Schwarz et al. 2014; Grandillo et al. 2013; Celik et al. 2017; Piosik
73 et al. 2019; Wang et al. 2020).

74 *S. pimpinellifolium* played a vital role in the development of heat-tolerant tomato varieties in
75 Taiwan (Ebert and Chou 2015). In Nigeria, it showed better responses to high humidity
76 compared to the cultivated elite tomatoes (Atugwu and Uguru 2012). Among many benefits of
77 gene introgression from *S. pimpinellifolium* is the increased fruit number, which is a major
78 contributory attribute to yield, in addition to resilience to high humidity (Wang et al. 2020).
79 These inherent benefits in *S. pimpinellifolium* is the motivation in selecting one of its lines
80 ‘LA2093’ to improve tomato for increased yield and adaptability to the humid tropical climes.

81 Evaluation of different population means performance of derived progenies at early segregating
82 generations as an approach to improve most economic traits especially yield and fruit quality
83 traits for selection have recently been reported in tomato (Finzi et al. 2020; Gomes et al. 2021).
84 Selection of any of such populations for improvement would be dependent on performance of the
85 populations across the multiple traits. However, a major challenge has been the difficulty of
86 knowing which statistical tool, preferably multivariate, is most suitable for evaluating multiplex
87 traits in either germplasm or breeding populations of crops.

88 Genotype selection or population improvement based on multiplex traits raises a lot of concern
89 in plant breeding. Genotype by yield*trait (GYT) biplot analysis is a novel tool with high
90 efficiency compared to other statistical tools (Yan and Frégeau-Reid 2018). The GYT biplot
91 positions genotypes or populations in hierarchy based on their levels in yield combination with
92 other target traits whether polygenic, monogenic or oligogenic other than yield, and expresses
93 trait profiles, which is their strengths and weaknesses. This is as opposed to the existing methods
94 which only concentrate efforts on genotype evaluation by their levels in individual traits
95 (Mohammadi 2019).

96 The objectives of this research are; (i) to assess the phenotypic variation in yield enhancing traits
97 in tomato using segregating lines to identify novel sources of improved performance, and (ii)
98 select population (s) among F₃ and BC₁F₂ populations, based on yield and other target traits
99 using GYT biplots analyses in a humid environment.

100 **2. Materials and Methods**

101 **2.1 Experimental sites description**

102 The present study was conducted at research field of the Department of Horticulture and Plant
103 Sciences, Jimma University, Ethiopia during the 2020/2021 and 2021/2022 cropping seasons.
104 Jimma is classified as warm to cold-environment locally known as “Weyna Dega” (Abdela et al.
105 2017), suitable for agriculture, with high degree of humidity, located on latitude 07°4’N,
106 longitude 36°50’E and altitude 1,710 m above the sea level in the south west, Oromia region of
107 Ethiopia. The monthly weather conditions of Jimma during the experiment seasons are presented
108 in Figures 1 and 2. Jimma is mainly covered with black, gray and red colored plastic clay soils
109 (Alemineh et al. 2020).

110 **2.2 Genetic materials**

111 A total of 276 potential breeding lines generated from four parental lines - elite parents of
112 cultivated tomato (*S. lycopersicum*) and wild parent ‘LA2093’ of *S. pimpinellifolium* from
113 California’s C.M. Rick Tomato Genetic Resource Center were used for this trial. The
114 domesticated cultivars were morphologically diverse in terms of superiority and weakness in the
115 traits studied. The four elite lines - ‘CLN2498D’ (D) and ‘CLN2714H’ (H) from Ethiopia and
116 ‘UC Dan INDIA’ (U) and ‘Tima’ (T) from Nigeria were used.

117 'CLN2498D' and 'UC Dan INDIA' were superior performing parents based on fruit yield and
118 shape/size of fruits (round and large) but had fewer number of fruits and susceptible of the humid
119 condition of the growing environment whereas 'CLN2714H' and 'Tima' were poor performers
120 in terms of fruit number production, fruit yield, having elongated medium fruit size, and longer
121 floral and fruit phenology. The wild parent (W) 'LA2093' was commonly used as pollen donor
122 to the four cultivated materials (females) using a bi-parental mating design by hand pollination
123 according to Ozores-Hampton (2014). According to Wang et al. (2020) 'LA2093' has prolonged
124 shelf life, an early flowering and maturing variety, quantitative disease resistant and relatively
125 resilient to humid environmental condition. It possesses enormous genetic potentials, having
126 desirable alleles for breeding choice tomato cultivars with appreciable economic qualitative and
127 quantitative traits performance.

128 Four F_1 crosses, *viz.*, 'H \times W', 'T \times W', 'D \times W', and 'U \times W' were obtained. Then two
129 different crosses were done. Firstly, the F_1 materials realized from each cross were crossed back
130 to the recurrent parents to obtain BC_1 . The BC_1 was also selfed to obtain BC_1F_2 and $BC_1F_{2:3}$
131 progenies, and secondly, the F_1 population selfed to produce F_2 . The F_2 of each cross was selfed
132 to produce F_3 . A total of eight populations were developed: four F_3 populations and four BC_1F_2
133 populations. The following populations were coded as presented in Table 1.

134 **2.3 Field management and experimental design**

135 The plant materials were evaluated using an alpha lattice experimental design with two
136 replicates. Each replicate contained 19 blocks whereas each block contained 15 plots. The
137 tomato seeds were first planted in plastic seed trays filled with sterilized top soil mixed with well
138 cured poultry manures and river sand in the ratio of 3: 2: 1, respectively by volume with the use

139 of a head pan. At 24 days after seedling emergence with the appearance of 4-5 true leaves,
140 uniform vigorous seedlings were transplanted to the field.

141 Each plot occupied by a particular genotype of each cross whether $F_{3:4}$ or $BC_1F_{2:3}$ consisted of
142 three rows of plant (30 plants/ plot) in an area of 1.5m × 5m with 0.5m × 0.5m inter-intra row
143 spacing. A distance of 1m and 0.5m alleys were ensured between blocks and plots, respectively.
144 Poultry droppings at the rate of 10 metric tons per hectare were worked into the soil within each
145 replicate 16 days before seedlings were transplanted. Cultural practices such as weed control,
146 irrigation, fertilizer (DAP-Di Ammonium Phosphate), fungicide (Ridomil-Mancozeb and
147 Metalaxyl-M), staking, pruning, and insecticide (Karate-Lambda-Cyhalothrin 5% EC) were
148 applied as suggested by Osei et al. (2010).

149 **2.4 Data collection**

150 Data were taken on 10 plants of each genotype in each replicate all from inner plants of each plot
151 eluding border plants. The following morphological traits were measured and recorded at 9 week
152 after transplanting during which the genotypes were expected to have completed vegetative
153 process as recommended by AVRDC guideline (Dinssa et al. 2015).

154 Plant height (PH cm) was measured from the plant base to the shoot tip. Number of leaves (NL),
155 primary branches (NBp), secondary branches (NBs), and nodes (NN) were counted.
156 Phenological traits included; number of days to first anthesis (DFA), 50% anthesis (D50A), first
157 fruit emergence (DFFE), 50% fruit set (D50FS), and first fruit ripening (DFFR) were taken from
158 the day of transplantation of seedlings. Fruit shelf-life traits included: number of days to first
159 fruit spoilage (D1stFSp) and 100% fruit spoilage (D100FSp) stored under room temperature
160 according to Arah et al. (2015).

161 The room temperature ranged from 18°C to 23°C while the relative humidity was from 84% to
162 91% throughout the shelf life duration. Total number of flowers per plant (TNFIPP) and total
163 number of fruits per plant (TNFrPP) were counted. Fruits were cut crosswise to count number of
164 locules per fruit (NLPF). The total number of mature fruits showing ripening initiation at second
165 harvest was weighed with electronic weighing balance and the fruit weight per plant (FWPP g)
166 recorded. The total fruit yield per hectare (TFYPH t/h) was estimated as described by Dinssa et
167 al. (2015).

168 **2.5 Statistical analysis**

169 Before computing the analysis of variance, test for homogeneity of residual variances was
170 carried out using F test where larger variance was divided by the smaller variance between the
171 two seasons, which suggested the two sets of data could be combined. Each population data
172 along parents was subjected to two-way analysis of variance (ANOVA) using R software (R
173 Core Team 2017) to analyze variance components.

174 Descriptive statistics and genetic parameters were done using the package “variability” in R as
175 recommended by Raj et al. (2020). The descriptive statistics were means, and ranges (minimum
176 and maximum), while the genetic parameters included genetic advance as percentage of mean
177 (GAM), genotypic coefficient of variation (GCV), phenotypic coefficient of variation (PCV);
178 broad sense heritability (h_{bs}) which was estimated as the ratio of genotypic variance (GV) to
179 phenotypic variance (PV). The GCV and PCV values were categorized as high (>20%), medium
180 (10–20%) and low (<10%) based on the recommendation of Burton and DeVane (1953). The
181 expected GAM (%) on 5% selection intensity were ranked as low (1 – 10%), intermediate (10 –
182 20%) and high (>20%) whereas broad sense heritability was categorized as low (0 – 30%),
183 intermediate (30 – 60%) and high (>60%) according to Khan et al. (2021). Comparison among

184 populations was based on these classified genetic parameters: GCV, PCV, h_{bs} and GAM using
185 graphs.

186 Data were further analyzed by the ‘genotype by yield*traits (GYT)’ biplots for the ‘Tester
187 Vector’ view showing relationship among yield-trait combinations, the ‘Average Tester
188 Coordination’ view ranking the genotypes based on their overall superiority and their strengths
189 and weaknesses, and ‘which wins where’ view or the polygon view highlighting genotypes with
190 outstanding profiles based on the genotype by yield*trait combined data, an option of GGE
191 biplot software version 6.3 (Yan 2001). We adopted the principle of GYT biplot analysis on the
192 various populations with ‘each population mean’ representing genotype of the GYT biplot
193 analysis.

194 From the original data (Supplemental Table S1), a GYT table was derived (Supplemental Table
195 S2), in which each column was a yield*trait combination. The method indicates that each trait
196 should either be multiplied or divided by total fruit yield according to the objectives of breeding.
197 To develop the GYT table, traits with favorable or desirable increased values (such as PH, NL,
198 NBp, NBs, NN, TNFIPP, TNFrPP, FWPP, D1stFSp, D100FSp, and NLPF) were multiplied with
199 yield using the multiplication operator (“*”) whereas traits with favorable decreased values (the
200 phenological traits) were used to divide yield using the division operator (“/”).

201 By this method, in the GYT table a larger value is always more desirable. A mean superiority
202 index (MSI) which involved all yield*trait combinations was also calculated based on the
203 standardized GYT table as described by (Yan and Frégeau-Reid 2018) using the formula as
204 follows:

205
$$P_{ij} = \frac{T_{ij} - \bar{T}_j}{S_j}$$

206 Where,

207 P_{ij} is the standardized value of genotype i for yield-trait combination j in the standardized table,
208 \bar{T}_{ij} is the original value of genotype i for yield-trait combination j in the GYT table
209 (Supplemental Tables S2), T_j is the mean across genotypes for yield-trait combination j , and S_j is
210 the standard deviation for yield-trait combination j . The biplots of GYT were based on singular
211 value decomposition of trait-standardized data (“Scaling = 1, Centering = 2”) and trait-focused
212 singular value partition (“SVP = 2”) according to Yan and Tinker (2005), for all the views
213 employed in the present trial except the ‘Average Tester Coordination’ view which showed SVP
214 = 1. Traits were regarded as ‘tester’ when using ‘relation among testers’ option. The ‘Tester
215 Vector’ view of the GYT biplots showed associations among the yield-trait combinations. The
216 ‘Average Tester Coordination’ view of the GYT biplots ranked the populations based on their
217 overall superiority and their strengths and weaknesses. The ‘which-won-where’ view of the GYT
218 biplots was used to highlight population with outstanding profiles.

219 Hierarchical clustering on principal components (HCPC) was done to group the genotypes based
220 on the measured traits, and the results were visualized using the *fviz_cluster* functions of the R
221 package “*factoextra*” for factor map (Kassambara and Mundt 2020).

222 **3. Results**

223 **3.1 Genetic variability, heritability and genetic advance for quantitative traits**

224 Despite the variations recorded in some weather elements for the two years, the test for
225 homogeneity carried out on the tomato traits data suggested the performance of a combined
226 analysis of variance. The result showed that genotypes within each population differed
227 significantly for total fruit yield and other traits studied ($P < 0.001$). The descriptive and genetic

228 parameters including h_{bs} and GAM of the traits in each population are summarized in
229 Supplemental Tables S4a & S4b to S7a & S7b.

230 Comparing the parental lines, 'CLN2498D' was the highest for NLPF (7.22), FWPP (3093.2 g)
231 and yield (64.03 t/h), followed by 'UC Dan INDIA' (6.48, 2955.1 g and 61.17 t/h, respectively)
232 while the wild parent, 'LA2093' displayed the highest values for PH (160.06 cm), NL (191.48),
233 NBp (12.53), NBs (45.72), NN (39.51), TNFIPP (511.78) and TNFrPP (496.25), highest days to
234 observe the fruit firmness and expressed the least days to observed all the floral and fruit
235 phenological traits. Variation of the populations from the better performing parent was also
236 shown in this study. For instance, 'pop_1_W/H1' showed higher maximum values for PH
237 (176.29 cm), TNFIPP (568.66), TNFrPP (534.15) and NLPF (7.35); 'pop_7_W/U1' for NL
238 (195.96), NBp (14.53), NBs (47.81), and NN (43.34); 'pop_8_W/U2' for D1stFSp (27.24 days)
239 and D100FSp (37.23 days) than those recorded in the better parents depending on trait.

240 'pop_5_W/T1' had the highest maximum value among the 8 populations for FWPP (2292.55g)
241 and fruit yield (46.15 t/h), the least maximum values for DFFE (28.23 days), and DFFR (48.82
242 days); 'Pop_7_W/U1' for DFA (15.93 days), D50A (20.04 days) and D50FS (36.81 days)
243 although the values were poorer than the better performing parents. Moreover, the mean values
244 showed similar results trend with the maximum range descriptions except that among the
245 populations, least D50FS (34.89 days) was recorded in 'pop_5_W/T1'.

246 All the 8 populations displayed low (<10%) GCV and PCV for the phenological traits: DFA,
247 D50A, DFFE, D50FS and DFFR. High (>20%) GCV and PCV were shown in all the populations
248 for PH. For NL, majority of the populations expressed medium to high GCV and PCV with the
249 exception of 'pop_4_W/D2' and 'pop_8_W/U2'. Intermediate (10-20%) or high (>20%) GCV

250 and PCV were recorded for FWPP and NLPF in all populations except ‘pop_5_W/T1’ which
251 showed low values for FWPP. All the populations displayed medium or high GCV and PCV
252 values for NBp except ‘pop_8_W/U2’, whereas for NBs all the populations were low apart from
253 ‘pop_1_W/H1’, ‘pop_5_W/T1’ and ‘pop_6_W/T2’.

254 ‘Pop_3_W/D1’ and ‘pop_7_W/U1’ showed high GCV and PCV for NN; leaving the rest
255 populations intermediate, whereas ‘pop_6_W/T2’ had low values. ‘Pop_3_W/D1’,
256 ‘pop_4_W/D2’, ‘pop_7_W/U1’ showed intermediate GCV and PCV values for TNFIPP and
257 TNFrPP while ‘pop_5_W/T1’ and ‘pop_6_W/T2’ could display similar category of GCV and
258 PCV only for TNFrPP. For the two traits which tested fruit durability after harvest, D1stFSp and
259 D100FSp, all the populations expressed medium or high GCV and PCV values with the
260 exclusion of ‘pop_3_W/D1’ and ‘pop_4_W/D2’, although ‘pop_7_W/U1’ displayed medium
261 GCV and PCV only for D100FSp. For fruit yield, ‘pop_2_W/H2’, ‘pop_3_W/D1’ and
262 ‘pop_7_W/U1’ displayed moderate GCV and PCV values. The difference between the GCV and
263 PCV values were negligible for all the traits in all populations.

264 All the populations exhibited very high magnitude of GV and PV for traits such as PH, NL,
265 TNFIPP, TNFrPP, and FWPP. Among the populations, ‘pop_1_W/H1’ displayed in magnitude
266 the highest GV and PV values for PH and NL; ‘pop_4_W/D2’ for TNFIPP and TNFrPP; and
267 ‘pop_6_W/T2’ for FWPP. The variations that existed between the PV and GV for all the traits in
268 all the populations were minimal.

269 High (>20%) GAM was displayed by all the populations for PH, NL, NBp, NN, and NLPF,
270 except ‘pop_4_W/D2’ for NL (13.13%); ‘pop_6_W/T2’ for NN (16.20%) and ‘pop_8_W/U2’
271 for NL (13.18%) and NBp (15.42%) which showed intermediate values. For NBs, high GAM

272 were expressed by 'pop_1_W/H1', 'pop_5_W/T1' and 'pop_6_W/T2'; intermediate by
273 'pop_2_W/H2', 'pop_3_W/D1' and 'pop_7_W/U1'. GAM values for DFA and D50A were low
274 (<10%) in 'pop_1_W/H1', 'pop_4_W/D2' and 'pop_6_W/T2' and moderate (10-20%) in
275 'pop_3_W/D1' and 'pop_8_W/U2'. However, 'pop_2_W/H2' displayed moderate value for
276 DFA whereas 'pop_5_W/T1' and 'pop_7_W/U1' exhibited similar class of GAM for D50A.

277 The expression of high GAM was observed in 'pop_3_W/D1', 'pop_4_W/D2', and
278 'pop_7_W/U1' for TNFIPP and TNFrPP leaving the rest with medium values for similar traits.
279 All the populations showed low (<10%) GAM for traits such as: DFFE, D50FS, and DFFR
280 except 'pop_6_W/T2' which displayed a weak intermediate value for DFFE. Apart from
281 'pop_5_W/T1' which expressed moderate GAM for FWPP, the rest populations showed high
282 values with 'pop_3_W/D1' being the highest. For D1stFSp and D100FSp, high GAM were noted
283 in 'pop_1_W/H1', 'pop_2_W/H2', 'pop_5_W/T1' and 'pop_6_W/T2'; intermediate values were
284 observed in 'pop_3_W/D1' and 'pop_4_W/D2'; while 'pop_7_W/U1' displayed high GAM for
285 D100FSp and moderate for D1stFSp. All the populations evaluated showed medium GAM for
286 total fruit yield except 'pop_2_W/H2', 'pop_3_W/D1' and 'pop_7_W/U1' which displayed high
287 values for the same trait.

288 High (>60%) broad sense heritability were found in all the populations for virtually all the traits
289 studied. However, moderate (30-60%) heritability estimates were consistently observed for
290 DFFR in all the populations except 'pop_6_W/T2' (63.20%) and 'pop_5_W/T1' (26.25%) which
291 showed high and low (<30%) values, respectively for similar trait. 'Pop_4_W/D2' expressed
292 intermediate heritability for DFFE (52.03%) and low value for D50FS (25.97%) leaving the rest
293 populations with high values for the same traits. Moderate heritability estimate was recorded in
294 'Pop_6_W/T2' for D50A (50.50%) among all the populations. Figures 3 to 7 present graphs

295 which compared the 8 populations for each quantitative trait and concentrated only on the
296 categorized genetic parameters such as genotypic coefficient of variation, phenotypic coefficient
297 of variation, heritability estimates in broad sense and genetic advance as a percentage of mean.

298 **3.2 Genotype by yield*trait (GYT) biplots using different views**

299 The idea behind the involvement of different views of the GYT biplot was to allow the data to be
300 investigated from different angles for better understanding of the relationship between
301 populations and yield*trait combinations, superior population(s) as well as the yield*trait profiles
302 of the populations. The total variation revealed by the PC1 and PC2 among the yield*traits
303 combinations of these views/models was 97.5%.

304 **3.2.1 Relationships among yield*trait combinations using the Tester Vector view of the** 305 **GYT biplot**

306 According to the Tester Vector view of the GYT biplot for the 8 populations, all yield*trait
307 interactions had a positive correlation with each other as shown by the acute angles between their
308 vectors (Figure 8). This may be due to the involvement of yield as a component of all yield*trait
309 combinations. This vital feature of GYT biplot makes it unique when compared to GT biplot as
310 genotypes or populations are easily ranked graphically and meaningfully based on their
311 yield*trait combinations.

312 The result showed that the magnitudes of angles among yield*D1stFSp, yield*D100FSp and the
313 rest yield*trait combinations were higher although all yield*trait combinations indicated high
314 positive correlation. Among all populations, 'pop_5_W/T1' and 'pop_1_W/H1' had the largest
315 values for yield*PH, yield*NN, yield*TNFIPP and yield*TNFrPP, although they had high
316 proximity with several other yield*trait combinations.

317 **3.2.2 Superiority vs. “weaknesses and strengths” of genotypes using the Average Tester**
318 **Coordination view of the GYT biplot**

319 Figure 9 represents the Average Tester Coordination view for the 8 tomato populations. This
320 view of the GYT biplot showed the superiority ranking of the populations based on their
321 yield*trait combinations. The single arrow line which passed through the biplot origin and the
322 average yield*trait interactions is the average tester coordinate (ATC) whereas the small circle
323 on the ATC showed the placement of the “average yield*trait combination,” which was
324 determined by the coordinates of all yield*trait combinations in the biplot. The double arrow
325 blue line is the general population mean. It divided the plot into two parts which separated the
326 populations based on their performance. The populations found on the side of the ATC arrow
327 (right) showed better performance for the surrounding yield*traits combinations whereas those
328 found at the opposite side (left) showed weak performance.

329 The best ranked population based on the yield*trait combinations was ‘pop_5_W/T1’ and was
330 followed by ‘pop_1_W/H1’, ‘pop_4_W/D2’, and ‘pop_6_W/T2’. In contrast, ‘pop_2_W/H2’,
331 ‘pop_7_W/U1’, ‘pop_3_W/D1’, and ‘pop_8_W/U2’ were ranked the poorest based on their
332 performance below the population mean for all the yield*trait combinations. Based on traits
333 profiles of the populations, ‘pop_5_W/T1’, ‘pop_1_W/H1’, and ‘pop_6_W/T2’ appeared to be
334 balanced for various yield*trait combinations; while ‘pop_4_W/D2’ was strong in the two shelf-
335 life traits: D1stFSp and D100FSp in combination with yield but weak in TNFIPP, TNFrPP, PH,
336 and NN. Despite populations general superiority or weakness stand, those placed below the ATC
337 such as ‘pop_5_W/T1’, ‘pop_1_W/H1’, ‘pop_6_W/T2’, and ‘pop_2_W/H2’ tended to have
338 relatively good levels of the traits below the ATC while having poor levels of the traits above the

339 ATC. The opposite is true for populations ('pop_7_W/U1', 'pop_3_W/D1', 'pop_8_W/U2' and
340 'pop_4_W/D2') placed above the ATC.

341 The weakness and strength of each population is presented in Supplemental Table S3. The best
342 populations were 'pop_5_W/T1' and 'pop_1_W/H1' based on the magnitude of their mean
343 values as expressed in the mean superiority index (MSI) table, in that order. Of these,
344 'pop_5_W/T1' did not have any negative value for all yield*traits combinations, while the
345 second placed 'pop_1_W/H1' exhibited negative values for D1stFSp. Apart from these best
346 performer populations mentioned which had positive MSI, the rest populations expressed
347 negative values for MSI.

348 **3.2.3 Genotypes performance on yield*trait combinations using which-won-where view of** 349 **the GYT biplot**

350 The result showed that 'pop_5_W/T1' accompanied by 'pop_1_W/H1' exhibited the best
351 performance for a majority of the yield*trait combinations (Figure 10). The only yield*trait
352 combination left out was yield*D1stFSp which projected 'pop_4_W/D2' as the best performer.
353 However, 'pop_4_W/D2' and 'pop_5_W/T1' shared equal but leading performance for
354 yield*D100FSp which appeared on the radiate line between the two populations. The result
355 showed that these populations mentioned were the best in combining fruit yield with the other
356 traits as observed in the present biplot.

357 **3.3 Clustering pattern analysis for the quantitative traits in F₃ and BC₁F₂ populations**

358 Clustering pattern analysis using hierarchical clustering on principal components (HCPC)
359 classified the lines into four clusters in the two population types (Figures 11 and 13). However,
360 transgressive segregation was observed for the studied traits in both F₃ and BC₁F₂.

361 There was significant structuring among the lines observed in F₃ composite populations. Among
362 F₃ populations (Figure 11), clusters D and U which encompassed originally lines from crosses
363 between ‘CLN2498D × LA2093’ and ‘UC Dan INDIA × LA2093’ contained transgressive or
364 overlapping lines (70.58% of all D lines) and (54.05% of all U lines), respectively from the two
365 populations. These two populations/clusters were characterized by less vigorous plants with
366 lower performance in virtually all traits compared to clusters H and T (Figure 12; Supplemental
367 Tables S4a, S5a, S6a, & S7a). Of the 37 lines of ‘UC Dan INDIA × LA2093’ cross (cluster U),
368 two lines ‘0210U1’ and ‘0211U1’ were lone genotypes (outliers) whereas ‘0171T1’ was the only
369 lone genotype from ‘Tima × LA2093’ cross (cluster T ‘pop_5_W/T1’).

370 Similar trend was found between clusters H and T of ‘CLN2417H × LA2093’ and ‘Tima ×
371 LA2093’ crosses, respectively. 8.57% of all H lines and 30.30% of all T lines were found
372 overlapping between clusters/populations H and T. Lines such as ‘0020H1’ and ‘0035H1’ of
373 cluster H found their way expressly among lines of cluster T whereas ‘0144T1’ of cluster T
374 showed slight drifting into cluster H. The cluster description is found in Figure 11.

375 The mean of each F₃ population plotted in a boxplot analysis displayed significant differences for
376 all the studied traits (Figure 12). Cluster H ‘pop_1_W/H1’ had the highest mean values for PH,
377 TNFIPP, and TNFrPP; while cluster T ‘pop_5_W/T1’ had the highest for NB_prim, NB_sec,
378 NN, FWPP, and fruit yield, the least DFFE, DFFR, also taking the shortest days to 100% fruits
379 shriveling. Number of leaves, D1stFSp, and D100FSp implicated cluster/population U
380 ‘pop_7_W/U1’ as the cluster with the highest mean values while having the least mean values for
381 DFA and D50A. In contrast, Clusters D ‘pop_3_W/D1’ and U consistently showed lower
382 performance comparatively for traits such as PH, NB_prim, NB_sec, NN, TNFIPP, TNFrPP,
383 including fruit yield.

384 With respect to BC₁F₂ populations (Figure 13), lots of transgressive segregations were also
385 observed. Three clusters, D, T and H showed profuse overlapping *viz.*, clusters D and T, as well
386 as clusters T and H. 32.35% of the 34 original lines of cluster D and 48.48% of all T lines
387 overlapped between clusters D and T, while 18.18% of all T lines and 8.82% of all H lines were
388 found transgressive between clusters T ‘pop_6_W/T2’ and H ‘pop_2_W/H2’. Very negligible
389 number of cluster U lines was found overlapped with cluster T. Apart from all the phenological
390 traits, NBp and NBs where cluster U performed best, cluster T took leadership for the rest yield
391 enhancing traits including yield itself (Supplemental Tables S4b, S5b, S6b, & S7b). There was
392 significant population structuring among the lines observed in BC₁F₂ composite population used
393 in this study.

394 Some lines were found elsewhere outside their original clusters. For instance, lines ‘0195T2’ and
395 ‘0194T2’ of cluster T were found in clusters H and U, respectively; line ‘0055H2’ of cluster H
396 drifted into cluster T, line ‘0124D2’ of cluster D fell into cluster H and ‘0272U2’ of cluster U
397 was found in cluster D. BC₁F₂ lines were generated through backcross to the recurrent parents.
398 The description of the clusters D, H, T, and U of the BC₁F₂ population is found in Figure 12.

399 Using the boxplot analysis, the mean of each BC₁F₂ cluster group showed significant differences
400 ($p < 0.05$) for all studied yield enhancing traits (Figure 14). Cluster H outperformed the rest
401 clusters in mean values for PH, TNFIPP, and TNFrPP, although it took the longest days to
402 achieve first anthesis, 50% anthesis, and 50% fruit set. Cluster U was consistent with the least
403 mean values for DFA, D50A, D50_FS, and DFFE, except DFFR where cluster T showed the
404 least mean value. Cluster U ‘pop_8_W/U2’ also had highest mean value for NB_sec. Cluster T
405 ‘pop_6_W/T2’ had the highest mean values for NN, FWPP, NLPF, and fruit yield. Clusters D and
406 U were comparatively higher for NL, NB_prim, D1stFSp, and D100FSp.

407 4. Discussion

408 4.1 Comparative analysis of variation among the selfed and backcrossed populations

409 Analysis of variance showed significant variation for virtually all traits studied, as well as wider
410 ranges for most traits in all the 8 populations including the F₃ and BC₁F₂ population types. The
411 highest maximum range and mean values among populations for various traits gave an indication
412 that selection for PH, TNFIPP, TNFrPP, and NLPF will favor ‘pop_1_W/H1’; NL, NBp, NBs,
413 NN, least DFA, D50A and D50FS will enhance ‘Pop_7_W/U1’; FWPP and fruit yield, least
414 DFFE and DFFR will improve ‘pop_5_W/T1’ of selfed (F₃) populations, whereas selection for
415 increased D1stFSp and D100FSp will favor the backcross population ‘pop_8_W/U2’. Gomes et
416 al. (2021) reported the presence of variability among the BC₁F₃ populations of Santa Cruz type
417 dwarf tomato which supported the selection of tomatoes that stood out for agronomic and quality
418 traits. De Oliveira et al. (2021) in another study reported genetic dissimilarities among 19 BC₁F₃
419 populations evaluated for agronomic and fruit quality traits. In the present study, evaluating F₃
420 and BC₁F₂ populations simultaneously opened up opportunity for comparison, of which the
421 selfed populations showed better variability and yield trait increased performance. Atugwu and
422 Uguru (2012) reported enormous variation and fruit size increase among early recombinants of a
423 cross between cultivated species and *S. pimpinellifolium* accession. The high or medium to high
424 GCV and PCV displayed for a majority of traits in one population or the other showed greater
425 phenotypic and genotypic variability among the lines of the affected populations, hence, higher
426 strength of the affected traits to make progress in selection. The opposite is the case for traits that
427 expressed low GCV and PCV values. The narrow differences between PCV and GCV for all the
428 traits in all populations showed less influence of the environment on the expression of these
429 traits, suggesting the presence of high coefficient of additive genetic variance and hence,

430 indicated that direct selection would be effective and provides desirable output for improvement
431 (Saleem et al. 2013). Furthermore, it reflects the potential for these affected traits to respond to
432 selection pressure on affected segregating populations, a case of evolvability of the traits (Hill
433 2010). Evolvability expresses a trait's capacity to adapt to various selection pressures to which a
434 population is subject (Cheung 2020). Heritability and genetic advance simultaneously give a
435 more reliable gain from selection in a breeding program. High or moderate to high heritability
436 followed by similar categories of genetic advance as percentage of mean for PH, NL, NBp,
437 NLPF, FWPP, fruit shelf life traits, and total fruit yield observed in one population or the other
438 indicated that the inheritance pattern in these traits were due to additive gene actions, which also
439 showed the effectiveness of selection to improve the traits at F_{2-3} generation (Henareh 2015).
440 The phenological traits *viz.*, DFA, D50A, DFFE, D50FS, and DFFR, which expressed low values
441 of genetic advance in a majority of the populations gave the traits a drawback for selection
442 although the heritability for some were high or moderate to high. Low GAM indicated the
443 involvement of non additive gene action in the inheritance of these traits, giving large scope for
444 heterosis breeding as an improvement approach (Saravanan et al. 2019). Saravanan et al. (2019)
445 reported high heritability combined with low genetic advance as a percentage of mean for days
446 to 50 % flowering in tomato. Among the populations studied, 'pop_1_W/H1' followed by
447 'pop_4_W/D2' and 'pop_6_W/T2' expressed the highest genetic divergence for PH, NL,
448 TNFIPP, TNFrPP, and FWPP based on their high magnitude of PV and GV. This revealed
449 options or signs of future success for obtaining best fruit producing, humid tolerant tomato lines.
450 Finzi et al. (2020) identified some tomato populations with the highest divergence and the most
451 promising for development of inbred lines with improved fruit traits among the 12

452 BC₁F₂ populations of dwarf round tomatoes. Genetic dissimilarity was also reported in BC₁F₃
453 populations of saladette tomato (De Oliveira et al. 2021).

454 **4.2 Tomato Selfed populations better than backcrossed populations**

455 The correlation coefficient between two particular traits is approximately equal to the cosine of
456 the angle across the space separating their vectors (Yan and Kang 2003). The positive correlation
457 witnessed virtually for all yield*trait combinations implied that yield combined with all the other
458 traits as indicated by the acute angles of their vectors in the biplot. Also, the long vector which
459 represents each of the yield*trait combination indicates very strong and high relationship among
460 the yield*trait combinations. These showed that one of the traits should be enough selection
461 criteria. A major advantage of relationship among yield*trait biplot view is its ability to identify
462 excessive traits, select fewer traits and reduce costs in traits measurement and management
463 without undermining experiment precision.

464 The trait profile experience showed excellent performance of selfed populations ‘pop_5_W/T1’
465 and ‘pop_1_W/H1’ for yield*TNFIPP, yield*TNFrPP, yield*PH and yield*NN, although they
466 had high proximity with the other yield*trait combinations. Hence, these populations were
467 considered promising, each with specific positive yield*traits associations. Relationship among
468 yield*trait biplot view is not considered as the best to check trait profiles of the genotypes (Yan
469 and Frégeau-Reid 2018). However, becoming aware of the interrelationships among the
470 yield*trait combinations make it easier to take a well informed decision on any genetic material
471 for breeding programs, aimed at selecting trait of interest for improvement (Mendonça et al.
472 2018; De Oliveira et al. 2019; Kendal 2019).

473 The Average Tester (trait) Coordination view of GYT biplot is effective as genotypes are ranked
474 on the account of their levels in combining yield with target traits, and simultaneously indicating
475 the strengths and weaknesses of the genotypes (Yan and Frégeau-Reid 2018). The results also
476 expressed the possibility of determining contrasting populations based on the yield*traits
477 combinations for improving tomato breeding. In the present study, ‘pop_5_W/T1’,
478 ‘pop_1_W/H1’, ‘pop_4_W/D2’, and ‘pop_6_W/T2 ranked as the best in combination of some
479 yield enhancing traits with fruit yield under cool tropical monsoon climatic conditions of Jimma,
480 Ethiopia, which showed genetic gain in tomato breeding program. This was based on the
481 magnitude of their mean values as expressed in the mean superiority index (MSI) table, which
482 was evidence of the yield*trait profiles of the populations. Of these, ‘pop_5_W/T1’ did not have
483 any negative value for all traits, which showed that this population had the best characteristic
484 performance across multi-traits studied; while ‘pop_1_W/H1’ which followed ‘pop_5_W/T1’
485 based on MSI table exhibited negative value only for D1stFSp indicating that this population had
486 low value for this traits among the best performers. The MSI ranked the tomato populations by
487 means of all yield*traits studied, where high values of MSI showed the best performing
488 population (s). Other studies have used MSI ranking to identify top performers, like Yan and
489 Frégeau-Reid (2018) for Oat, Mohammadi (2019) for durum wheat, and Lance et al. (2020) for
490 red spring wheat.

491 Using the polygon model, ‘pop_5_W/T1’ and ‘pop_1_W/H1’ also exhibited the best
492 performance for a majority of the yield*trait combinations which showed they were the best in
493 combining fruit yield with the rest traits in the humid ecology. ‘pop_4_W/D2’ together with
494 ‘pop_5_W/T1’ having taken the longest time to achieve the initial and 100% fruit spoilage
495 proved to possess more gene for fruit firmness from the wild parent ‘LA2093’ among the other

496 populations. The polygon view of genotype by yield*trait (GYT) biplot has recently been used to
497 study yield*trait combination profiles of genotypes, genotype evaluation and selection for
498 improvement in several crop species including oat (Yan and Frégeau-Reid, 2018; Yan et al.
499 2019), sesame (Boureima and Yaou 2019), durum wheat (Mohammadi 2019; Kendal 2019),
500 cowpea (De Oliveira et al. 2019; Araújo et al. 2021), peanut (Mahmoud et al. 2020), cotton
501 (Peixoto et al. 2022), and *Jatropha curcas* (Purwati et al. 2022).

502 **4.3 Clustering pattern analysis**

503 The clustering pattern was supported by the significant differences observed among selfed and
504 backcrossed populations for all seventeen investigated traits. This variation registered among
505 these tomato populations/clusters may be due to differences and similarities in genetic
506 constitution of the genotypes. This is partly in agreement with the report of Kanneh et al. (2016b)
507 who noted that variation observed among tomato interspecific genotypes was due to differences
508 in genetic and environment conditions. Interspecific hybridization can mix genetic materials
509 from two different species, which has proven to be an effective way to increase
510 phenotypic variability (Wang et al. 2020). The significant structuring among the lines of F₃ and
511 BC₁F₂ composite populations although with a bit of transgressive segregation and overlapping
512 lines contradicts the findings of Yu et al. (2008) and Bajgain et al. (2016) who reported no
513 significant population structure among the recombinant inbred lines of the nested association
514 mapping population design. According to the authors, the lack of structure among the lines could
515 be credited to the parents, which were diverse from each other. However, the population
516 structure pattern observed in F₃ and BC₁F₂ composite populations in the present study followed
517 the relatedness found among the recurrent parents in an earlier genetic characterization study

518 (Ene et al. 2022). The closeness in relationship among the recurrent parents could have
519 contributed to the observed population structuring pattern among the lines.

520 The high percentage of overlapping lines observed between clusters D and U, as well as clusters
521 H and T for the studied traits in F₃ populations indicated a level of close tie between the
522 overlapping clusters. It could be suspected that the genetic distance between the parental lines
523 that generated lines of these clusters that showed overlapping are similar. Ene et al. (2022)
524 reported the grouping of the recurrent parents of clusters D (CLN2498D) and U (UC Dan
525 INDIA) in the same cluster while those of clusters H (CLN2417H) and T (Tima) in the same
526 cluster showing a level of relatedness. Although each cluster was of different interspecific cross,
527 they were half sib progenies sharing a common parent (pollen donor-wild parent 'LA2093') from
528 the initial cross. This established a level of relationship among the progenies altogether. Similar
529 implication could be attributed to the same lines overlapping or drifting witnessed between
530 clusters D and T, as well as clusters T and H of BC₁F₂ populations. Rieseberg et al. (1999)
531 reported that transgressive segregation is favored by high genetic distance between the parental
532 lines, preferably of different species.

533 The outliers '0210U1' and '0211U1' lines from cluster U; lines '0020H1' and '0035H1' of
534 cluster H which found their way among lines of cluster T; and '0144T1' of cluster T which
535 showed slight drifting into cluster H, revealed transgressive behavior of the segregating lines
536 among selfed populations. A situation where little minority of recombinants are outliers relative
537 to the range of parental phenotypes, and is commonly observed in plant breeding populations
538 (Boyle et al. 2017). Whereas this phenomenon has been linked to complementary action of gene
539 and epistasis, the biochemical, physiological, and molecular bases have not been fully
540 understood (Pabuayon et al. 2021). Interspecific transgressive individuals have been argued to

541 represent a potential source of novel genetic variation in crop species as they can potentially
542 affect characters of adaptive significance (De Vicente and Tanksley 1993; Singh et al. 2018a;
543 Pabuayon et al. 2021).

544 Similar implication could be likened to the lines '0195T2' and '0194T2' of cluster T which
545 found their way into clusters H and U, respectively; line '0055H2' of cluster H which drifted into
546 cluster T, line '0124D2' of cluster D which fell into cluster H and '0272U2' of cluster U finding
547 its way into cluster D, among BC₁F₂ populations. De Vicente and Tanksley (1993) reported that
548 interspecific transgressive lines of tomato possessed characteristics that allowed them to occupy
549 new ecological niches where others could not or better competed in existing environments.
550 Epistatic interactions of parental alleles and complementary action of additive alleles are
551 regarded as being responsible for the superior or inferior attributes of transgressive segregants
552 (Dittrich-Reed and Fitzpatrick 2013).

553 Among selfed (F₃) populations, desirable performance appeared for PH, TNFIPP, and TNFrPP in
554 cluster H (pop_1_W/H1); NB_prim, NB_sec, NN, FWPP, fruit yield, early DFFE and DFFR in
555 cluster T ('pop_5_W/T1'); and NL, D1stFSp, and D100FSp in cluster U ('pop_7_W/U1'). For
556 BC₁F₂, desirable performance was observed in cluster U ('pop_8_W/U2') for early DFA, D50A,
557 D50_FS, and DFFE; cluster T (pop_6_W/T2) for NN, FWPP, NLPF, and fruit yield; clusters D
558 and U for NL, NB_prim, D1stFSp, and D100FSp. The findings suggested that improvement of
559 these traits through single plant selection would favor the genotypes of the respective clusters
560 and could be beneficial for future tomato breeding program, after verifying their consistencies
561 over different environments (Gonzalo et al. 2020). It further advocated that high yielding and
562 adaptable lines to high humidity could emerge from interspecific hybridization following single
563 plant selection (Gaur et al. 2008).

564 **5. Conclusions**

565 Genetic variability study revealed considerable dissimilarity among the lines within each
566 population and between populations that would be useful for fruit yield improvement and
567 adaptability to tropical humid climates. High or moderate to high heritability and high genetic
568 advance as percentage of mean for PH, NL, NB_prim, NN, NLPF, TNFIPP, TNFrPP, FWPP,
569 D1stFSp, D100FSp, and total fruit yield noticed in one population or the other indicated that
570 their inheritance pattern were due to additive gene actions, which suggested selection as an
571 effective improvement approach. Among the F₃ and BC₁F₂ populations, ‘pop_1_W/H1’,
572 ‘pop_4_W/D2’ and ‘pop_6_W/T2’ which expressed the highest genetic divergence for PH, NL,
573 TNFIPP, TNFrPP, and FWPP were most promising for development of inbred lines with
574 improved fruit traits. Selection for increased post-harvest durability will favor ‘pop_8_W/U2’.

575 Application of GYT biplots models in this study presents a novel approach to tomato population
576 improvement based on multiplex traits. ‘pop_5_W/T1’ and ‘pop_1_W/H1’ of F₃ population,
577 ranked the best in combining some yield enhancing traits with fruit yield, indicating genetic gain
578 and showing adaptability to the growing environment. GYT biplots could help to conquer the
579 general challenge of crop selection on multiple traits which has been a problem in plant breeding.

580 Clustering pattern showed significant differences among F₃ and BC₁F₂ populations for all studied
581 traits. Cluster T (‘pop_5_W/T1’) among F₃ populations, and cluster T (pop_6_W/T2) among
582 BC₁F₂ populations showed desirable performance for fruit weight and fruit yield, and of course
583 any improvement program for fruit yield under humid condition would favor the genotypes of
584 these clusters. Using multivariate analysis, transgressive segregants ‘0210U1’, ‘0211U1’, and

585 '0171T1' of selfed (F₃) population were observed. They are believed to represent a potential
586 source of novel genetic variation for future tomato breeding program.

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592 **Data Availability Statement**

593 All data set generated during and/or analyzed to support the findings of this study are phenotypic
594 data and can be made available from the corresponding author on request.

595 **Competing Interests**

596 The authors declare that there are no conflicts of interest, financial or nonfinancial, directly or
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799 **Table 1:** Population naming description

No.	Code	Description/ pedigree
1.	Pop_1_W/H1 (F ₃ Cluster H)	F ₃ population developed by selfing F ₂ hybrids of a cross between ‘CLN2714H’ (H) and ‘LA2093’ (W)
2.	Pop_2_W/H2 (BC ₁ F ₂ Cluster H)	BC ₁ F ₂ population developed by selfing backcross to the recurrent parent (BC ₁) from a cross between ‘CLN2714H’ (H) and ‘LA2093’ (W); H = recurrent parent
3.	Pop_3_W/D1 (F ₃ Cluster D)	F ₃ population developed by selfing F ₂ hybrids of a cross between ‘CLN2498D’ (D) and ‘LA2093’ (W)
4.	Pop_4_W/D2 (BC ₁ F ₂ Cluster D)	BC ₁ F ₂ population developed by selfing backcross to the recurrent parent (BC ₁) from a cross between ‘CLN2498D’ (D) and ‘LA2093’ (W); D = recurrent parent

5.	Pop_5_W/T1 (F ₃ Cluster T)	F ₃ population developed by selfing F ₂ hybrids of a cross between 'Tima' (T) and 'LA2093' (W)
6.	Pop_6_W/T2 (BC ₁ F ₂ Cluster T)	BC ₁ F ₂ population developed by selfing backcross to the recurrent parent (BC ₁) from a cross between 'Tima' (T) and 'LA2093' (W); T = recurrent parent
7.	Pop_7_W/U1 (F ₃ Cluster U)	F ₃ population developed by selfing F ₂ hybrids of a cross between 'UC Dan INDIA' (U) and 'LA2093' (W)
8.	Pop_8_W/U2 (BC ₁ F ₂ Cluster U)	BC ₁ F ₂ population developed by selfing backcross to the recurrent parent (BC ₁) from a cross between 'UC Dan INDIA' (U) and 'LA2093' (W); U = recurrent parent

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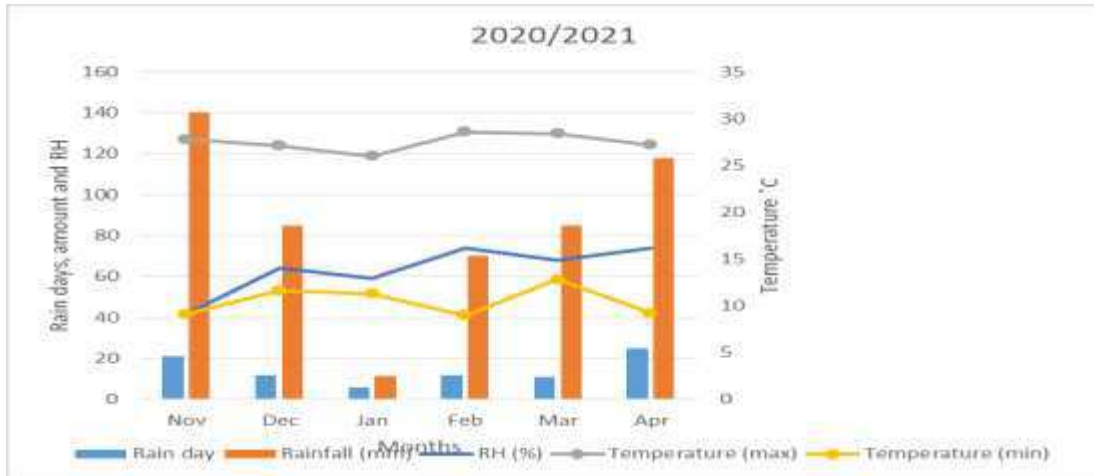
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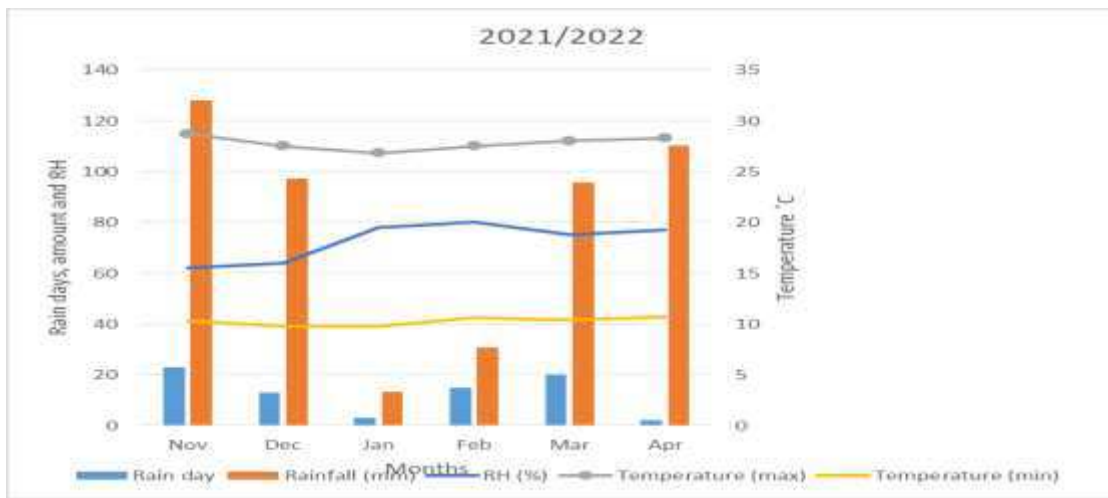
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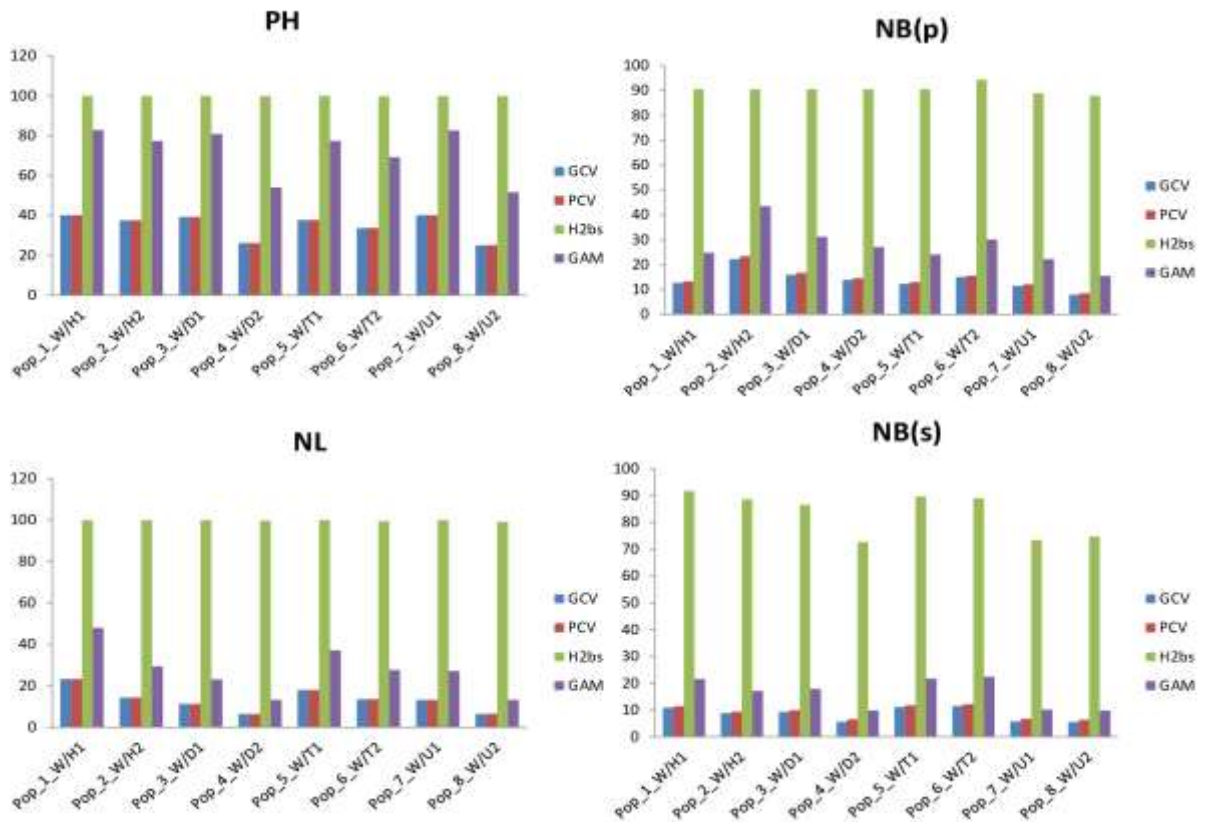
808 **Figure 1:** Monthly weather conditions of Jimma during the 2020/2021 experiment season



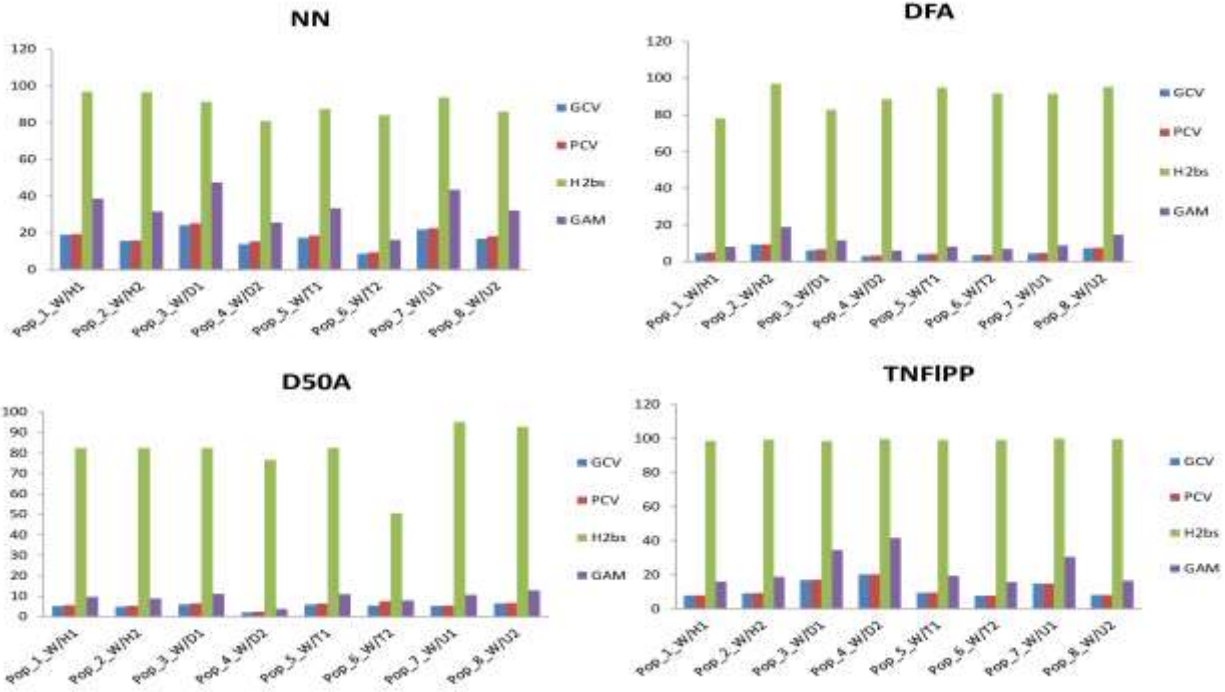
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810 **Figure 2:** Monthly weather conditions of Jimma during the 2021/2022 experiment season

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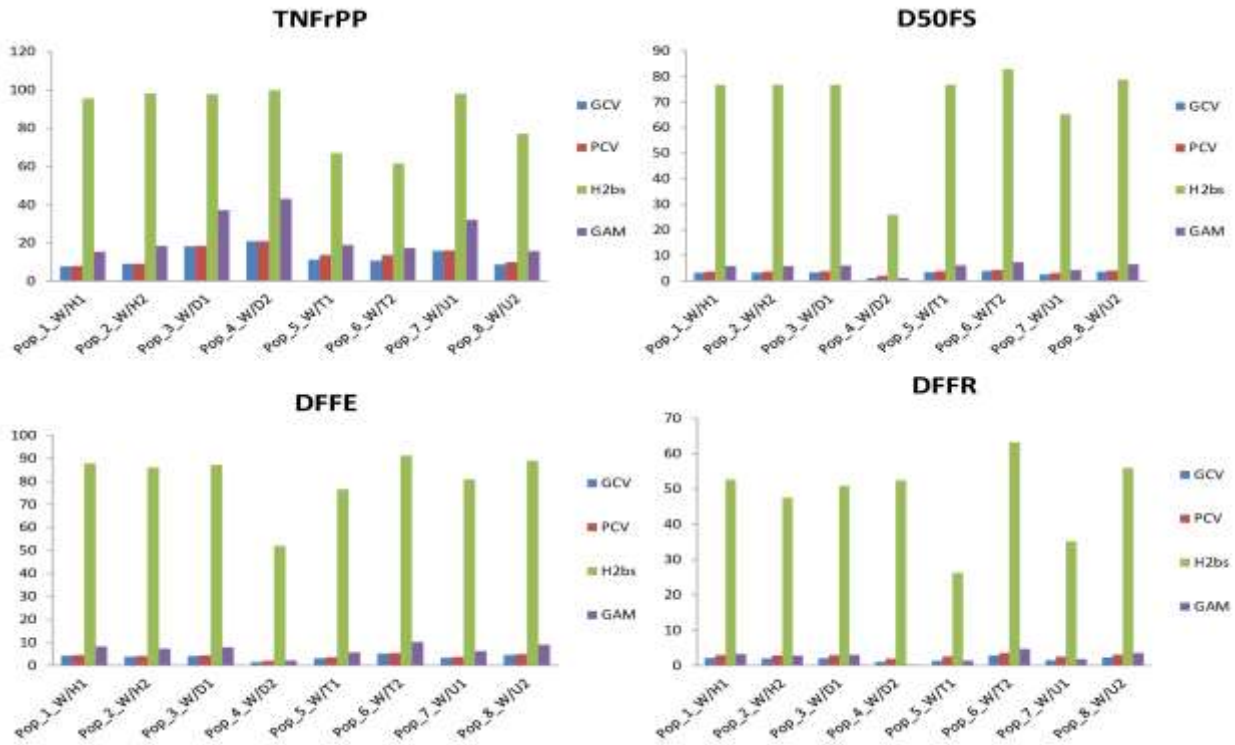


812 **Figure 3:** Genetic variation, broad sense heritability and genetic advance among tomato populations for
 813 plant height (PH), number of primary branches (NBp), number of leaves (NL) and number of secondary
 814 branches (NBs)



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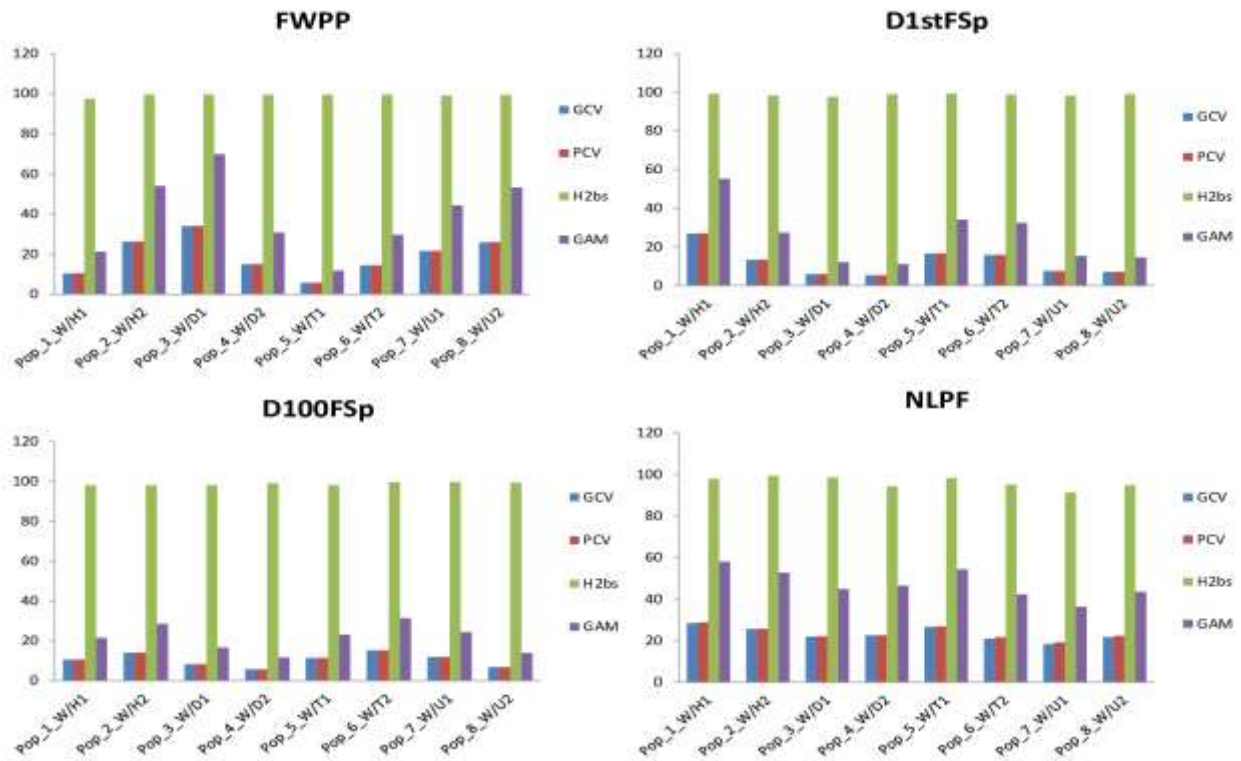
816 **Figure 4:** Genetic variation, broad sense heritability and genetic advance among tomato populations for
 817 number of nodes (NN), days to first anthesis (DFA), days to 50% anthesis (D50A) and total number of
 818 flower per plant (TNFIPP)



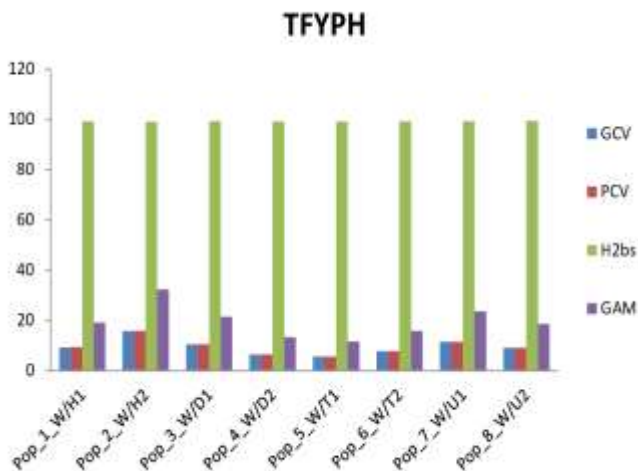
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820 **Figure 5:** Genetic variation, broad sense heritability and genetic advance among tomato populations for

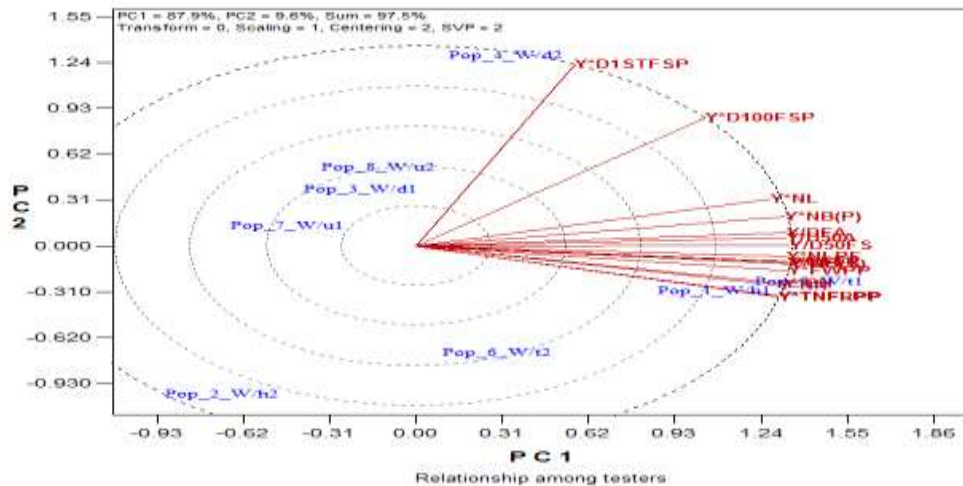
821 total number of fruit per plant (TNFrPP), days to 50% fruit set (D50FS), days to first fruit emergence
 822 (DFFE) and days to first fruit ripening (DFFR)



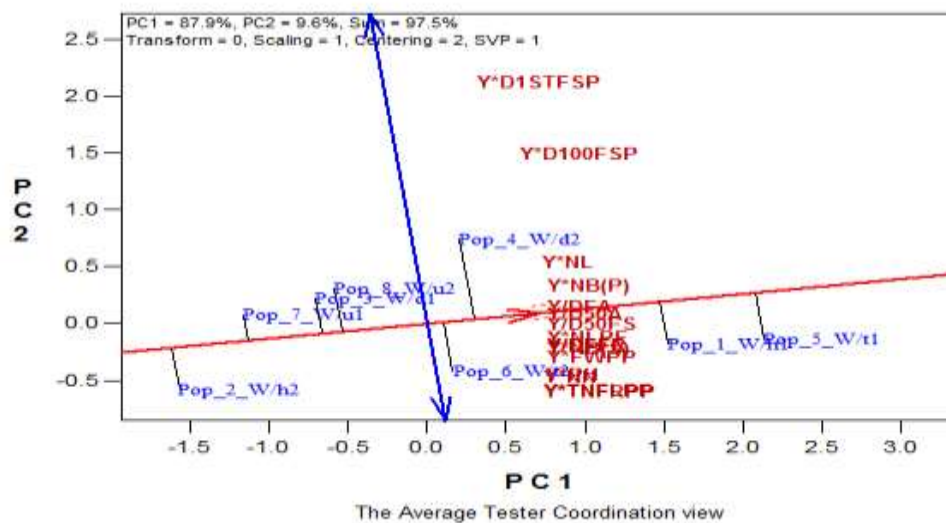
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 824 **Figure 6:** Genetic variation, broad sense heritability and genetic advance among tomato populations for
 825 fruit weight per plant (FWPP), days to first fruit spoilage (D1stFSp), days to 100% fruit spoilage
 826 (D100FSp) and number of locules per fruit (NLPF)



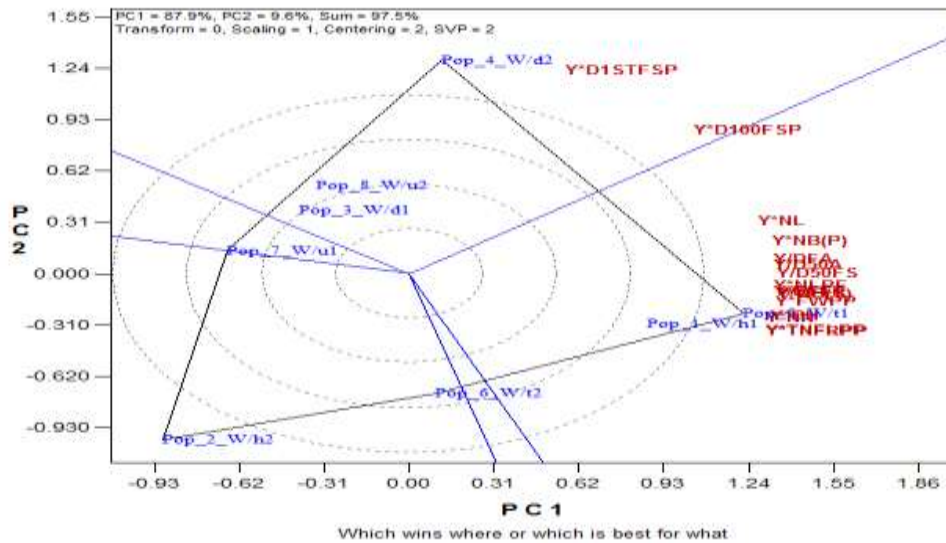
828 **Figure 7:** Genetic variation, broad sense heritability and genetic advance among tomato populations for
 829 total fruit yield per hectare



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 832 **Figures 8:** The Tester Vector view showing relationship among testers/traits of genotype by yield*trait (GYT) biplot of
 833 8 populations based on yield*traits combinations; PH (cm): Plant height, NL: Number of leaves, NB(p): Number of
 834 primary branches, NB(s): Number of secondary branches, NN: Number of nodes (at 9 week after transplanting), DFA:
 835 Days to first anthesis, D50A: Days to 50% anthesis, TNFIPP: Total number of flower per plant, TNFrPP: Total number
 836 of fruit per plant, FWPP (g): Fruit weight per plant, DFFE: Days to first fruit emergence, D50FS: Days to 50% fruit set,
 837 DFFR: Days to first fruit ripening, D1stFSp: Day to initial fruit spoilage after harvest, D100FSp: Days to 100% fruit
 838 spoilage after harvest, NLPF: Number of locule per fruit, Y (t/h): Total fruit yield per hectare, Pop_1_W/H1 (F₃ of 'H ×
 839 W'), Pop_2_W/H2 (BC₁F₂ of 'H × W'), Pop_3_W/D1 (F₃ of 'D × W'), Pop_4_W/D2 (BC₁F₂ of 'D × W'), Pop_5_W/T1
 840 (F₃ of 'T × W'), Pop_6_W/T2 (BC₁F₂ of 'T × W'), Pop_7_W/U1 (F₃ of 'U × W'), Pop_8_W/U2 (BC₁F₂ of 'U × W')

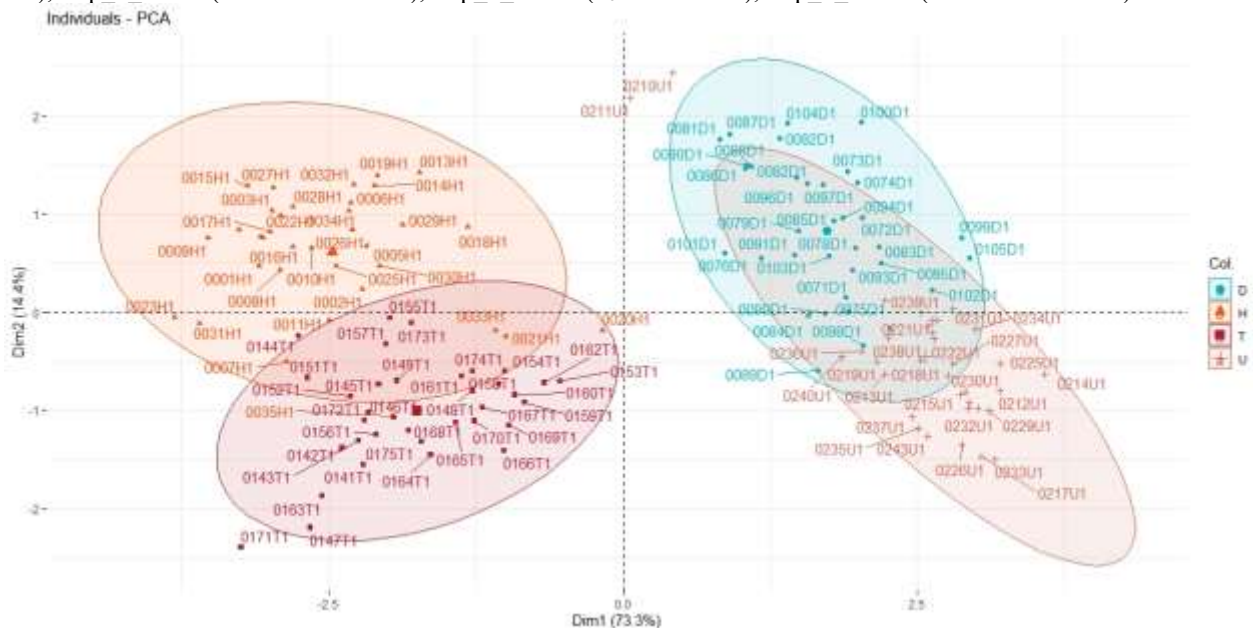


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 843 **Figures 9:** The Average Tester Coordination view of genotype by yield*trait (GYT) biplot involving 8 populations
 844 based on multi-trait; PH (cm): Plant height, NL: Number of leaves, NB(p): Number of primary branches, NB(s):
 845 Number of secondary branches, NN: Number of nodes (at 9 week after transplanting), DFA: Days to first anthesis,
 846 D50A: Days to 50% anthesis, TNFIPP: Total number of flower per plant, TNFrPP: Total number of fruit per plant,
 847 FWPP (g): Fruit weight per plant, DFFE: Days to first fruit emergence, D50FS: Days to 50% fruit set, DFFR: Days
 848 to first fruit ripening, D1stFSp: Day to initial fruit spoilage after harvest, D100FSp: Days to 100% fruit spoilage
 849 after harvest, NLPF: Number of locule per fruit, Y (t/h): Total fruit yield per hectare, Pop_1_W/H1 (F₃ of 'H ×
 850 W'), Pop_2_W/H2 (BC₁F₂ of 'H × W'), Pop_3_W/D1 (F₃ of 'D × W'), Pop_4_W/D2 (BC₁F₂ of 'D × W'), Pop_5_W/T1
 851 (F₃ of 'T × W'), Pop_6_W/T2 (BC₁F₂ of 'T × W'), Pop_7_W/U1 (F₃ of 'U × W'), Pop_8_W/U2 (BC₁F₂ of 'U ×
 852 W')



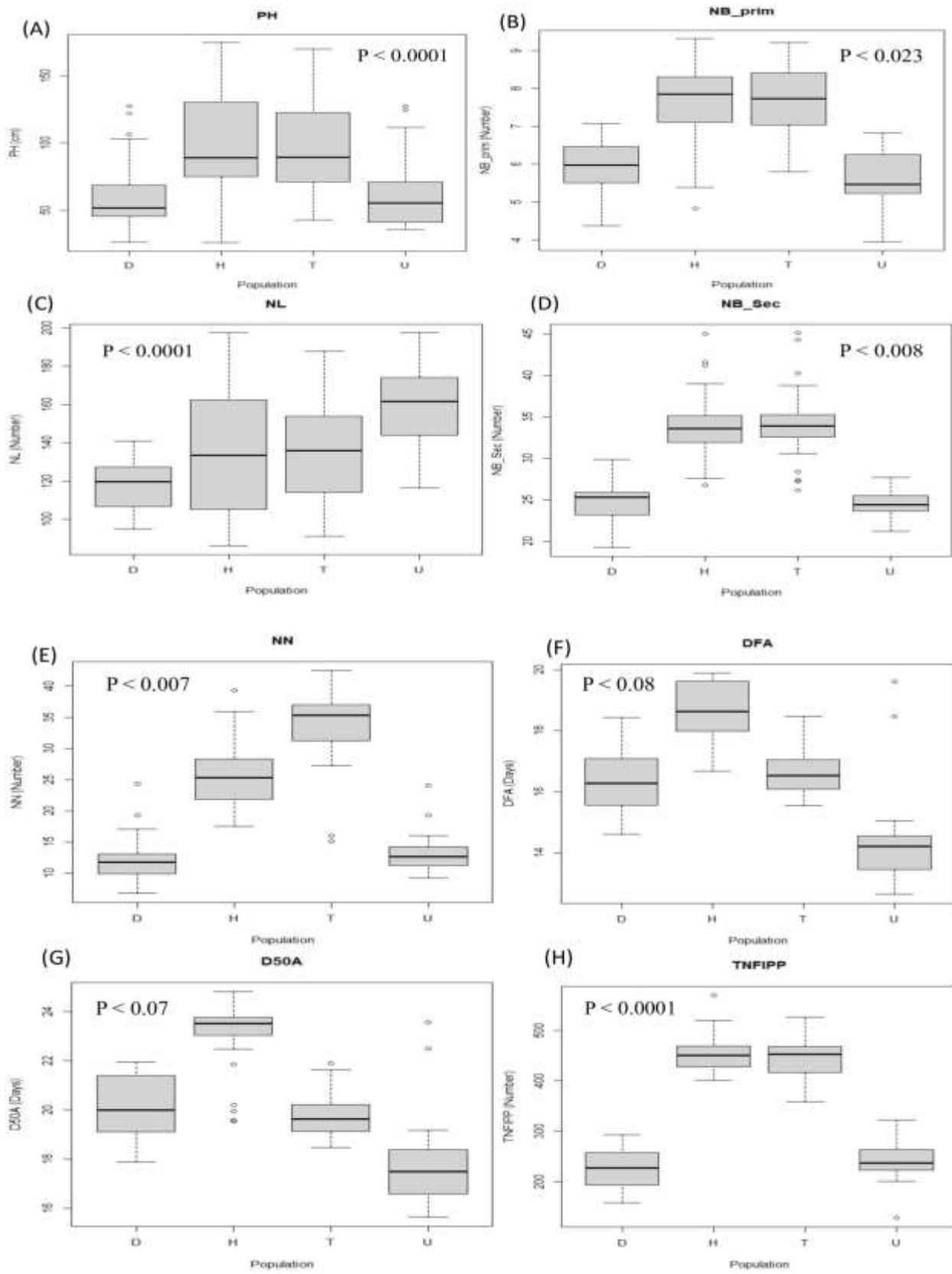
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Figures 10: The which-won-where view of genotype by yield*trait (GYT) biplot involving 8 populations based on multi-traits; PH (cm): Plant height, NL: Number of leaves, NB(p): Number of primary branches, NB(s): Number of secondary branches, NN: Number of nodes (at 9 week after transplanting), DFA: Days to first anthesis, D50A: Days to 50% anthesis, TNFIPP: Total number of flower per plant, TNFRPP: Total number of fruit per plant, FWPP (g): Fruit weight per plant, DFFE: Days to first fruit emergence, D50FS: Days to 50% fruit set, DFFR: Days to first fruit ripening, D1stFSp: Day to initial fruit spoilage after harvest, D100FSp: Days to 100% fruit spoilage after harvest, NLPF: Number of locule per fruit, Y (t/h): Total fruit yield per hectare, Pop_1_W/H1 (F₃ of 'H × W'), Pop_2_W/H2 (BC₁F₂ of 'H × W'), Pop_3_W/D1 (F₃ of 'D × W'), Pop_4_W/D2 (BC₁F₂ of 'D × W'), Pop_5_W/T1 (F₃ of 'T × W'), Pop_6_W/T2 (BC₁F₂ of 'T × W'), Pop_7_W/U1 (F₃ of 'U × W'), Pop_8_W/U2 (BC₁F₂ of 'U × W')



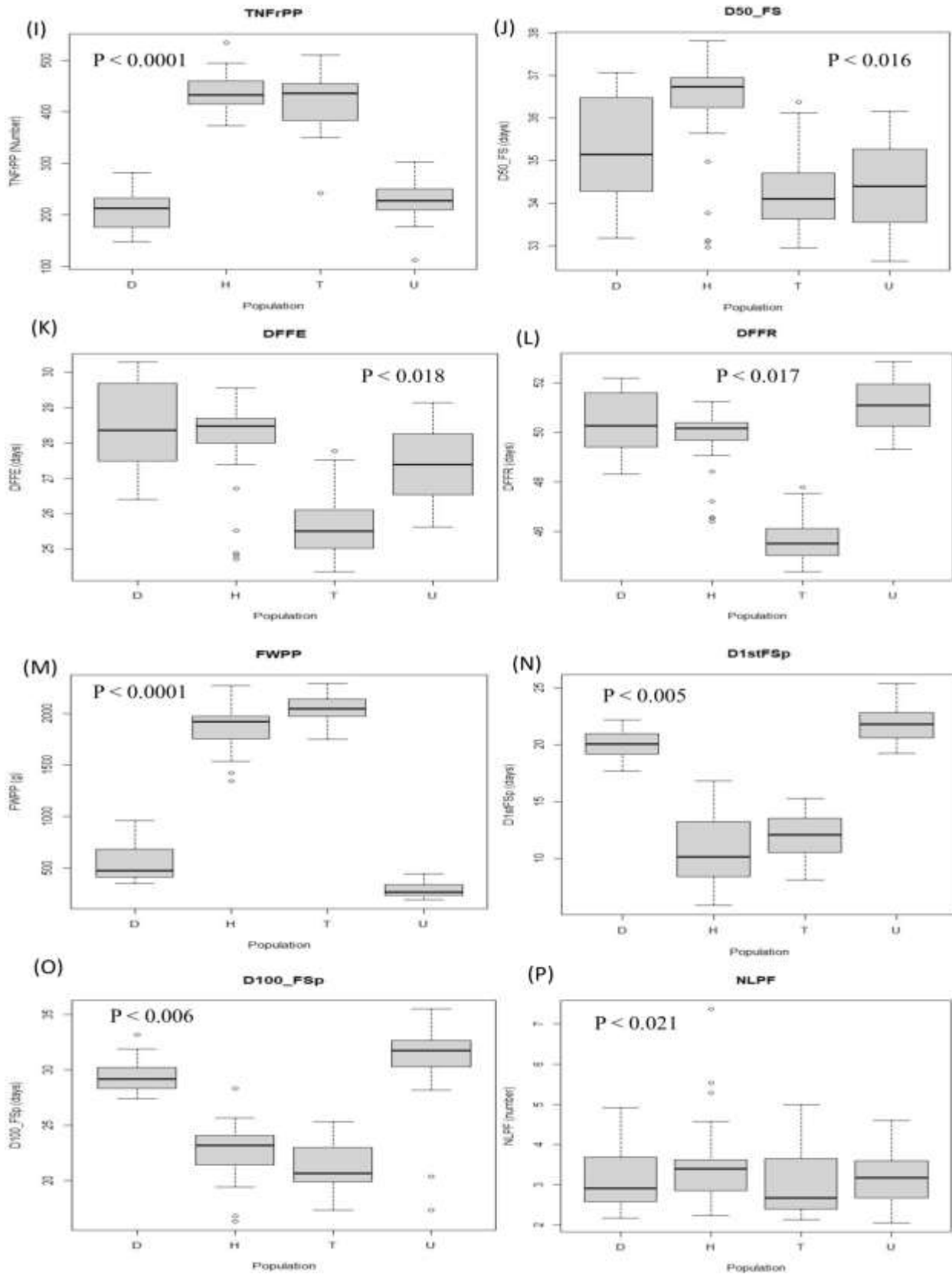
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Figure 11: Factor map showing the clustering pattern of 139 F₃ lines of 4 biparental populations of tomato based on the hierarchical clustering on principal components analysis (HCPC). Cluster D: 'pop_3_W/D1' (n = 34), Cluster H: 'pop_1_W/H1' (n = 35), Cluster T: 'pop_5_W/T1' (n = 33), and Cluster U: 'pop_7_W/U1' (n = 37).



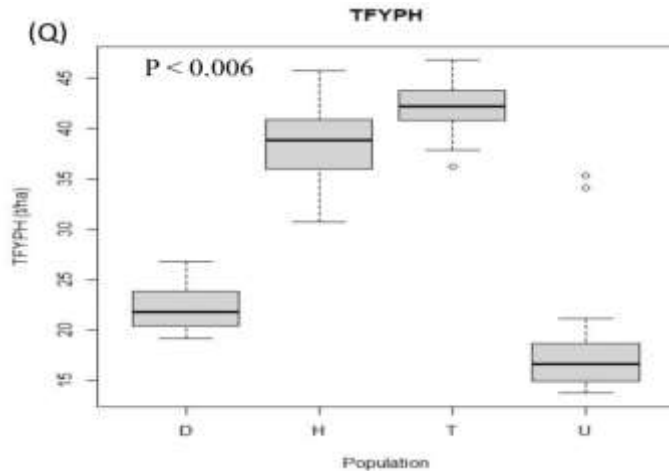
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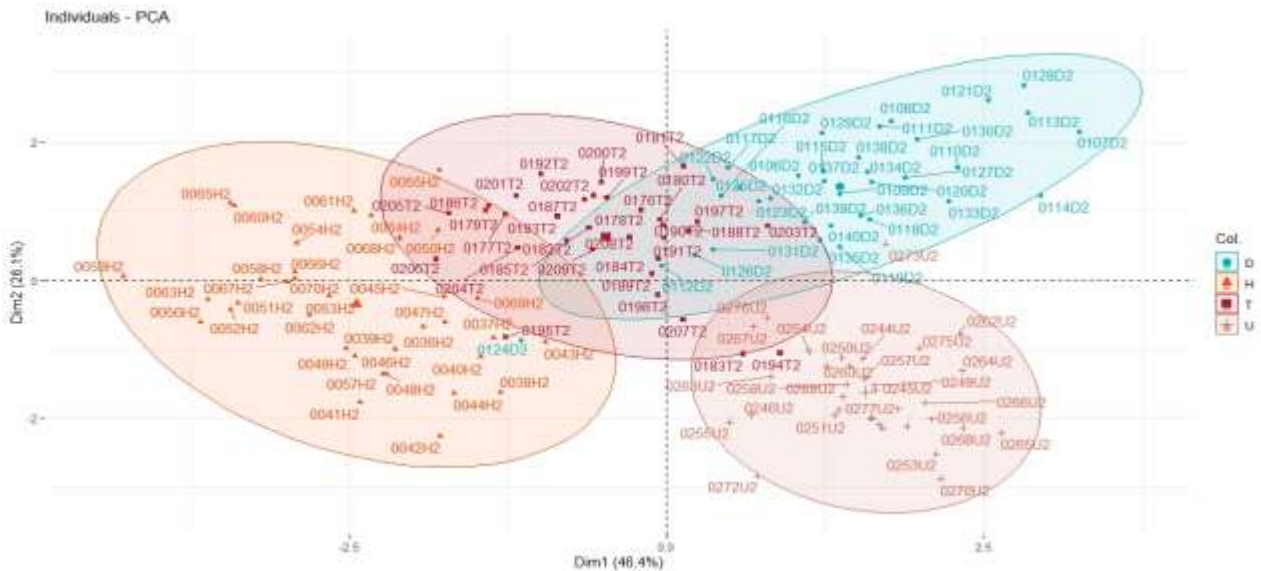
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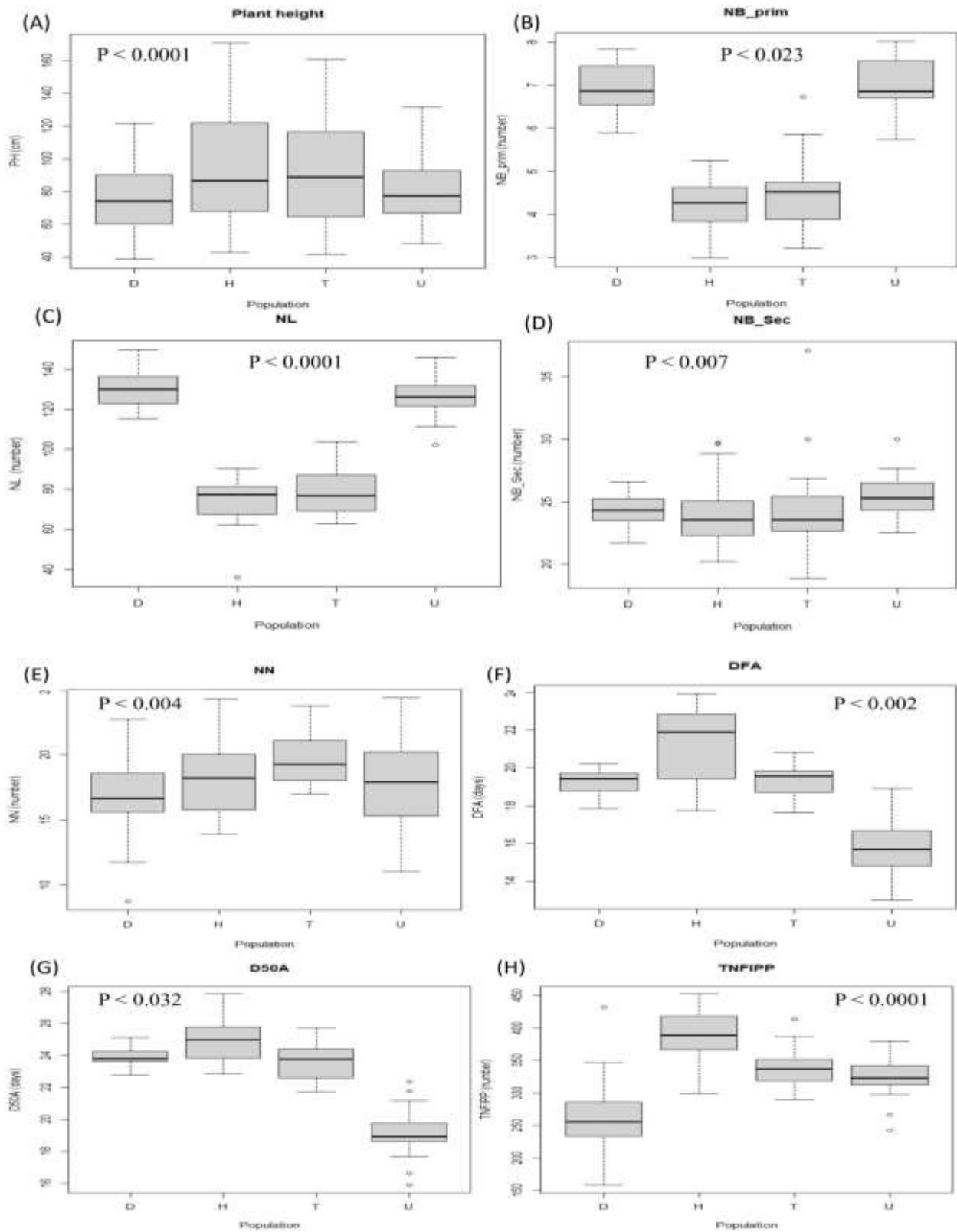


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 874 **Figure 12:** Phenotypic means for seventeen yield enhancing traits (A to Q) among the four F₃ clusters of
 875 139 segregating lines of tomato using box plots. (A) Plant height (cm). (B) Number of primary branches.
 876 (C) Number of leaves. (D) Number of secondary branches. (E) Number of nodes. (F) Days to first
 877 anthesis. (G) Days to 50% anthesis. (H) Total number of flowers per plant. (I) Total number of fruits per
 878 plant. (J) Days to 50% fruit set. (K) Days to first fruit emergence. (L) Days to first fruit ripening. (M)
 879 Fruit weight per plant (g). (N) Days to 1st fruit spoilage. (O) Days to 100% fruit spoilage. (P) Number of
 880 locules per fruit. (Q) Total fruit yield per hectare. Population D: ‘pop_3_W/D1’ (n = 34), Population H:
 881 ‘pop_1_W/H1’ (n = 35), Population T: ‘pop_5_W/T1’ (n = 33), and Population U: pop_7_W/U1’ (n =
 882 37).

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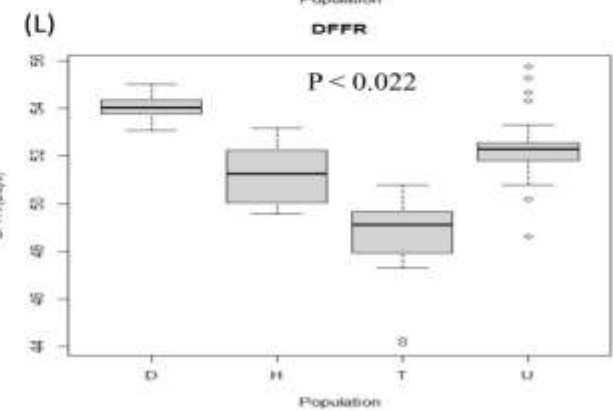
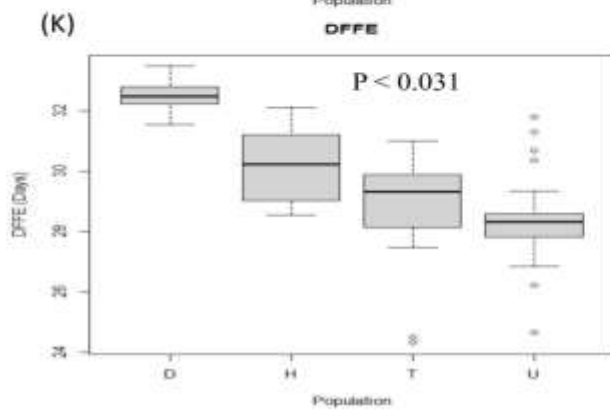
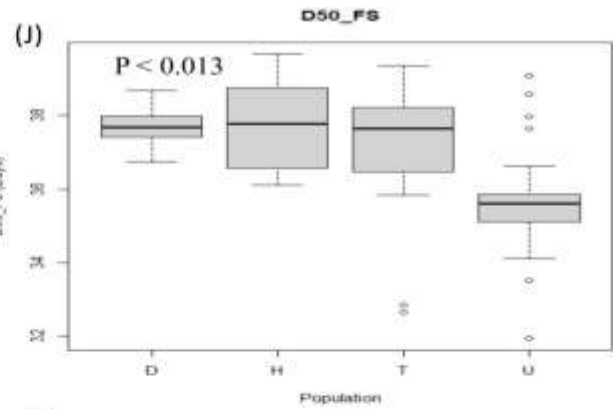
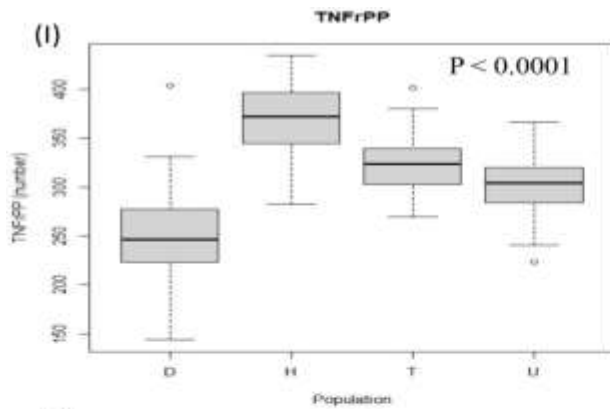


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 885 **Figure 13:** Factor map showing the clustering pattern of 137 BC₁F₂ lines of 4 biparental populations of
 886 tomato based on the hierarchical clustering on principal components analysis (HCPC). Cluster D:
 887 ‘pop_4_W/D2’ (n = 34), Cluster H: ‘pop_2_W/H2’ (n = 34), Cluster T: ‘pop_6_W/T2’ (n = 33), and
 888 Cluster U: ‘pop_8_W/U2’ (n = 36).

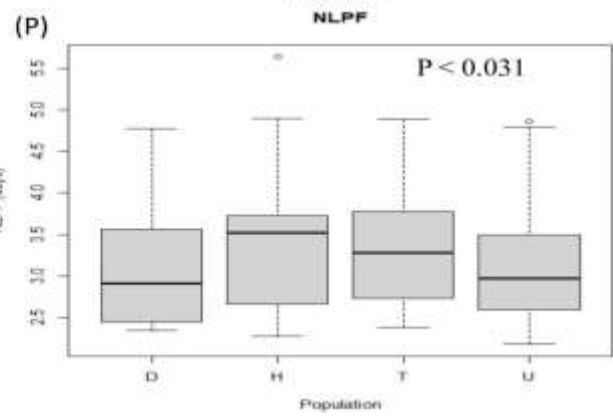
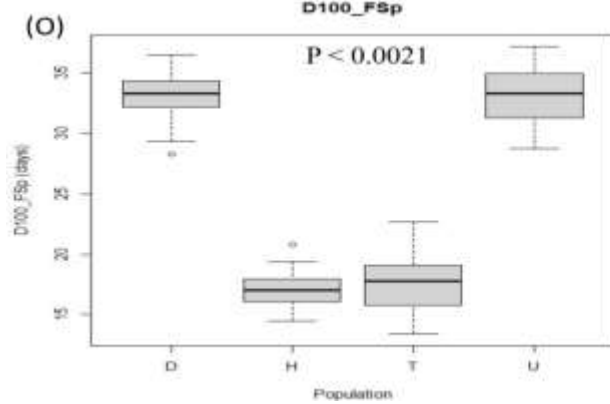
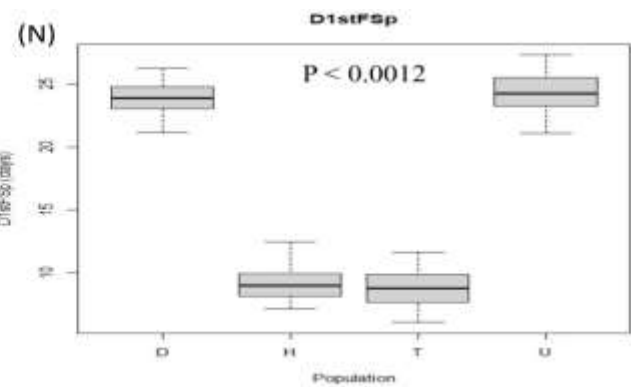
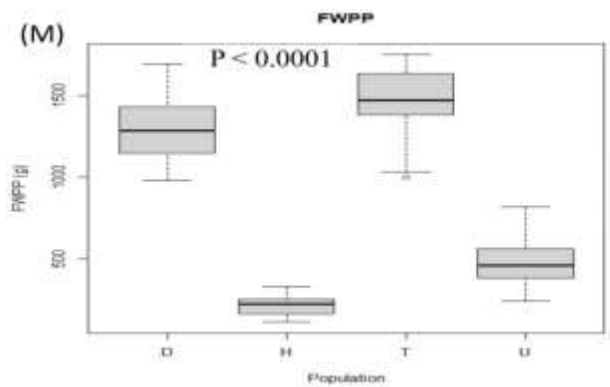


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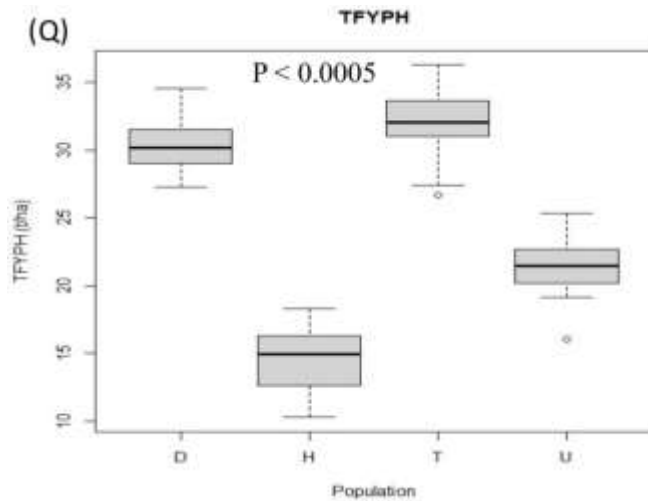
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 894 **Figure 14:** Phenotypic means for seventeen yield enhancing traits (A to Q) among the four BC₁F₂ clusters
 895 of 137 segregating lines of tomato using box plots. (A) Plant height (cm). (B) Number of primary
 896 branches. (C) Number of leaves. (D) Number of secondary branches. (E) Number of nodes. (F) Days to
 897 first anthesis. (G) Days to 50% anthesis. (H) Total number of flowers per plant. (I) Total number of fruits
 898 per plant. (J) Days to 50% fruit set. (K) Days to first fruit emergence. (L) Days to first fruit ripening. (M)
 899 Fruit weight per plant (g). (N) Days to 1st fruit spoilage. (O) Days to 100% fruit spoilage. (P) Number of
 900 locules per fruit. (Q) Total fruit yield per hectare. Population D: ‘pop_4_W/D2’ (n = 34), Population H:
 901 ‘pop_2_W/H2’ (n = 34), Population T: ‘pop_6_W/T2’ (n = 33), and Population U: ‘pop_8_W/U2’ (n =
 902 36)

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